The Asymmetric Synthesis of the [D₈]-Labeled (-)-(S)-Dihydroxyverbacine, the Terminal Precursor in the Biogenesis of the Macrobicyclic Spermine Alkaloids Aphelandrine and Orantine

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Dedicated to Professor Jürgen O. Metzger on the occasion of his 60th birthday

The asymmetric synthesis of the unlabeled and $[D_8]$ -labeled terminal precursors, **4** ((-)-(S)-dihydroxyverbacine) and **19**, respectively, in the biogenesis of the spermine alkaloids aphelandrine (**5**) and orantine (**6**), respectively, is described. A partial synthesis of the alkaloid (-)-(S)-[(E)-4-methoxycinnamoyl]buchnerine (**10**) is also presented.

Introduction. – The macrobicyclic spermine alkaloid aphelandrine (5) is found in the roots of *Encephalosphaera lasiandra* and, together with its diastereoisomer orantine (6), in several *Aphelandra* species. In addition, orantine was isolated from *Chaenorhinum minus*, *Schweinfurthia papilionacea* (Scrophulariaceae), and from *Ephedra* sp. (Ephedraceae) [1]. Both alkaloids contain a 13-membered and a 17-membered ring, with spermine and two *para*-coumaric acids as precursor moieties. The two hypotheses proposed for the biogenesis of these alkaloids are shown in *Scheme 1*. The first involves the N(1),N(5)-di(p-coumaroyl)spermine (1), which first undergoes cyclization by *Michael* addition to the compound (S)-4 and subsequently yields, *via* phenol coupling, aphelandrine (5) [2]. The second possibility is the transformation of monoacylated spermine derivative 2 by *Michael* addition to the macrocycle (S)-3, which is further acylated to (S)-4. Subsequent phenol coupling leads to aphelandrine (5) or orantine (6).

Compound 3 was recently detected in the roots of A. squarrosa and named prelandrine [3]. The presence of 3 in Aphelandra plants supports its possible key role in the biogenetic pathway of aphelandrine (5) and strongly favors the second pathway mentioned above [3]. Although the acylated derivative 4 of prelandrine (3) was not detected in the plant material, it seems to represent the terminal biogenetic precursor of aphelandrine (5) and orantine (6).

The last step in the biosynthesis of this type of alkaloids is proposed to be a diastereoselective intramolecular oxidative phenol coupling that leads to the substituted benzofuran ring of aphelandrine (5 (11S,17S,18S)) or orantine (6 (11S,17R,18R)). The presumable substrate for this diastereoselective intramolecular coupling is compound (S)-4, the dihydroxy derivative of the alkaloid (S)-verbacine, which was isolated earlier from different *Verbascum* species [4]. The (Z)-isomer of its (S)-di-O-methyl derivative has been isolated from *Clerodendrum buchneri* [5].

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Scheme 1

Since microsomal-bound cytochrome P-450 enzymes are known to participate in highly regio- and stereoselective phenol-coupling reactions in benzylisoquinoline alkaloids [6][7], it was expected that an enzyme of this type might be responsible for

the phenol coupling of the compound (S)-4 in the biosynthesis of aphelandrine (5) and orantine (6). To prove whether this type of enzyme, indeed, is responsible for the formation of the benzofuran ring of aphelandrine, a method for the isolation of a biochemically active microsomal fraction from the roots of A. squarrosa was developed. Additionally, highly sensitive analytical methods for the in vitro enzymatic studies were introduced [8].

To validate the hypothesis that (S)-dihydroxyverbacine (4) is the substrate for the enzymatic phenol coupling and thus the terminal precursor in the biosynthesis of aphelandrine (5) and orantine (6), the unlabeled and $[D_8]$ -labeled derivatives, 4 and 19, respectively, were synthesized. In the present paper, the syntheses of these two compounds for the *in vitro* enzymatic studies, and the synthesis of the alkaloid (-)-(S)-[(E)-4-methoxycinnamoyl]buchnerine (10) are described.

Results and Discussion. – Unlabeled (S)-dihydroxyverbacine (**4**) was synthesized in four steps starting from the synthetically prepared naturally occurring alkaloid (S)-buchnerine (**7**) [9] (Scheme 2). The bicyclic compound (S)-**8** was quantitatively obtained from (S)-**7** with 1 equiv. of HCHO. Additional acylation of (S)-**8** at N(9) gave the macrobicycle (S)-**9**, which was further hydrolyzed in the presence of NH₂OH (a HCHO interceptor) to compound (S)-**10**. The (Z)-isomer of (S)-**10**, (-)-(S)-[(Z)-4-methoxycinnamoyl]buchnerine, is a naturally occurring alkaloid detected earlier in *Clerodendrum buchneri* (Verbenaceae) [5]. O-Demethylation of (S)-**10** with BBr₃ led to (S)-dihydroxyverbacine (**4**).

The enzymatic assay with (S)-dihydroxyverbacine (4) showed the presence of aphelandrine (5) in the reaction medium, but it was impossible to distinguish between aphelandrine formed by the intramolecular cyclization of (S)-4 and the endogenous aphelandrine present in small amounts in the plant material (cell-microsomal fraction). To answer the question of whether (S)-dihydroxyverbacine ((S)-4) is indeed the precursor for the terminal diastereospecific macrocyclization, it was necessary to conduct experiments with labeled material.

The synthesis of the $[D_8]$ -(S)-dihydroxyverbacine (19) is depicted in *Scheme 3*. Compound (S)-11 was prepared according to the procedure for the asymmetric synthesis of the alkaloid (S)-buchnerine (7) [9]. The cyclization of (S)-11 was performed according to a modified *Richman-Atkins* protocol in DMF in presence of Cs_2CO_3 [10]: the $[D_8]$ -labeled butane-1,4-diyl moiety was introduced to (S)-11 (\to (S)-14), using (1,1,2,2,3,3,4,4- D_8)butane-1,4-diyl bis(methanesulfonate) (13), which was obtained from the commercially available [1,1,2,2,3,3,4,4- D_8]butane-1,4-diol (12). The electrochemical detosylation [11] of compound (S)-14 provided $[D_8]$ -(S)-buchnerine (15) in excellent yield. With 1 equiv. HCHO in MeOH, compound (S)-15 led quantitatively to the CH_2 -bridged compound (S)-16, which was additionally acylated at N(9) with (E)-3-(4-methoxyphenyl)prop-2-enoyl chloride to give (S)-17. By mild acid hydrolysis in the presence of NH_2OH , compound (S)-17 was converted quantitatively to (S)-N(1)-[(E)-4-methoxycinnamoyl][D_8]buchnerine ((S)-18). O-Demethylation of (S)-18 with BBr_3 gave the [D_8]-labeled precursor (S)-19.

The $[D_8]$ -labeled precursor (S)-19 can now be used as a substrate for the enzymatic oxidative phenol-coupling experiments with microsomal fractions from the roots of A. squarrosa. These biochemical experiments will pave the way to answer the question of

whether the proposed terminal precursor (S)-4 and its labeled analog (S)-19 are converted to aphelandrine or orantine, respectively, by a cytochrome P-450.

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Scheme 3

Experimental Part

General. [1,1,2,2,3,3,4,4-2H₈]butane-1,4-diol (98 atom-% D) was purchased from Aldrich. TLC: Merck precoated plates Kieselgel 60 F₂₅₄ were used, detection by UV (254 or 366 nm), Schlittler [12], and Dragendorff reagents. CC: Kieselgel 60 (70–230 mesh) from Merck. M.p.: Büchi 510 melting-point apparatus; uncorrected. Optical rotation: Perkin-Elmer 241 polarimeter. NMR: Bruker AC-300, ARX-300, or AMX-600 (¹H) and Bruker ARX-300 (75 MHz) or AMX-600 (150 MHz; ¹³C; only selected, well-defined signals are given); chemical shifts δ in ppm rel. to Me₄Si as internal standard. ESI-MS: Finnigan TSQ 700 mass spectrometer. (+)-(2S)-2-(4-Methoxyphenyl)-1,5,9,14-tetraazabicyclo[12.3.1]octadecan-4-one (8). To a soln. of (-)-(S)-buchnerine (7; 85 mg, 0.23 mmol) in 5 ml MeOH was added 37% aq. HCHO soln. (19 μl, 0.23 mmol). After several min, the mixture was evaporated, and the residue was dissolved in CHCl₃. The soln. was washed with dil. aq. NH₃ soln. and aq. sat. NaCl soln., dried (Na₂SO₄), and evaporated to give (S)-8 quantitatively. Colorless

glass-like solid. TLC (CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): R_f 0.74. $[a]_D^{20} = +6.27$ (c = 1.7, CHCl₃). ¹H-NMR (CDCl₃): 8.95 (br. s, CONH···N); 7.04, 7.03 (2d, J = 9, 2 arom. H); 6.84 (d, J = 9, 2 arom. H); 4.16–3.97 (br. m, PhCHN); 3.79 (s, MeO); 3.57–3.1 (m, 3 H); 3.07–1.82 (m, 14 H); 1.82–1.17 (m, 9 H). ¹³C-NMR (CDCl₃): 171.9 (C=O); 158.8 (arom. quat. C); 129.0 (arom. quat. C); 128.9, 113.3 (CH=CH); 63.5 (PhCN); 55.1 (MeO). ESI-MS: 374 ($[M+H]^+$).

(+)-(2S)-9-[(E)-3-(4-Methoxyphenyl)prop-2-enoyl]-2-(4-methoxyphenyl)-1,5,9,14-tetraazabicyclo[12.3.1]octadecan-4-one (9). To a suspension of 74 mg (0.29 mmol) of 1-methyl-2-chloropyridinium iodide [13] and 43 mg (0.24 mmol) of (*E*)-4-methoxycinnamic acid in a mixture of 2 ml of CH₂Cl₂ and 0.15 ml of Et₃N, cooled to 0°, was added dropwise a soln. of 90 mg (0.24 mmol) of (*S*)-8. The mixture was stirred in the dark at r.t. for 16 h. The mixture was diluted with CHCl₃, washed with 10% aq. soln. of K_2 CO₃ and H_2 O, dried (Na₂SO₄), and evaporated. The residue was dissolved in CHCl₃ and purified by CC (alumina, consecutively CHCl₃, AcOEt/MeOH 9:1) to obtain (*S*)-9 as colorless glass-like solid (109 mg, 85%). TLC (AcOEt/MeOH 4:1): R_f 0.45, $[a]_D^{20} = +5$ (c=2, CHCl₃). 1 H-NMR (CDCl₃): 9.55, 9.35 (2 br, s, CONH···N); 7.68, 7.67 (2d, J=15, PhCH=CHCO); 7.46 (d, J=9, 2 arom. H); 6.97 (d, J=9, 2 arom. H); 6.92 –6.8 (m, 4 arom. H); 6.69, 6.67 (2d, J=15, PhCH=CHCO); 3.97 (br. d, PhCHN); 3.83 (s, MeO); 3.8 (s, MeO); 3.75 –2.1 (m, 16 H); 1.98 –1.3 (m, 8 H). 13 C-NMR (CDCl₃): 172, 166 (2 C=O); 160.6, 159.1 (arom. quat. C); 142.2, 141.9, 129.2, 129, 128.1, 114.9, 114.1, 113.4 (C=C); 63.4 (PhCN); 55.2, 55.1 (2 MeO). ESI-MS: 535 ([M+H]+).

(−)-(8S)-1-[(E)-3-(4-Methoxyphenyl)prop-2-enoyl]-8-(4-methoxyphenyl)-1,5,9,13-tetraazacycloheptadecan-6-one (10). A mixture of 80 mg (0.15 mmol) of (S)-9 and 200 mg of NH₂OH · HCl in 5 ml of 1% aq. HCl soln. was heated at 70° for 1 h. The mixture was washed with CHCl₃, alkalinized with 25% aq. NH₃ soln., and extracted with CHCl₃. The org. extract was washed with H₂O, dried (Na₂SO₄), and evaporated to give 60 mg (77%) of (S)-(10). Colorless, glass-like solid. TLC (CHCl₃/MeOH/25% aq. NH₃ soln. 8:2:0.5): R_t 0.45. [α] $_0^2$ 0 = −12.6 (c = 1.5, CHCl₃). 1 H-NMR (CDCl₃, mixture of conformers): 8.46 (br. s, 0.5 H, CONH ··· N); 7.67 (d, J = 15, PhCH=CHCO); 7.48 (d, J = 8, 2 H); 7.35 (br. s, 0.5 H, CONH); 7.26 −7.09 (m, 2 arom. H); 6.93 −6.85 (m, 3 arom. H, H−C(8')); 6.76 −6.56 (m, 2 arom. H); 3.94 −3.91 (br. d, PhCHN); 3.84 −3.13 (m, 12 H, including 2 MeO at 3.84, 3.8, 3.79); 2.76 −2.32 (m, 8 H); 1.94 −1.43 (m, 10 H). 13 C-NMR (CDCl₃): 172, 166 (2 C=O); 161, 159, 142.4, 129.3, 127.9, 127.6, 115, 114.1, 113.8 (C=C); 58.9 (PhCN); 55.2, 55.1 (MeO). ESI-MS: 523 ([M + H] $^+$).

(-)-(8S)-1-[(E)-3-(4-Hydroxyphenyl)prop-2-enoyl]-8-(4-hydroxyphenyl)-1,5,9,13-tetraazacycloheptadecan-6-one Dihydrochloride ($\mathbf{4} \cdot 2$ HCl). To a soln. of (S)- $\mathbf{10}$ (60 mg, 0.11 mmol) in dry CH₂Cl₂ (5 ml), cooled to -78° (dry ice/acetone bath), 1.5 ml (1.5 mmol) of 1m CH₂Cl₂ soln. of BBr₃ were added. The mixture was kept at -78° for 1 h, then stirred for 16 h at r.t. The reaction was quenched by adding of 3 ml of 90% aq. MeOH soln. and evaporated to dryness *in vacuo* (50°). The excess BBr₃ was removed by repetitive addition of MeOH saturated with gas. HCl and evaporation. The solid residue was washed several times with CHCl₃, dissolved in CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:0.3 and purified by CC (consecutively CHCl₃, CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:0.3). The eluate was evaporated to dryness. The residual free base of (S)- $\mathbf{4}$ was transformed immediately to its dihydrochloride by addition of MeOH saturated with gas. HCl and evaporated to dryness: 50 mg (77° %) of (S)- $\mathbf{4}$ -2 HCl. Colorless solid. All steps were performed under red light to prevent (E) \rightarrow (Z) photoisomerization. TLC (CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): R_f 0.5. [α] $_D^{\circ}$ 0 = -1.5 (c = 1.3, MeOH). ¹H-NMR ((D₄)MeOH): 7.6-7.43 (m, including 2 arom. H and PhCH=CHCO, d, J = 15, at 7.55); 7.37, 7.36 (2d, J = 8, 2 arom. H); 6.93-6.84 (m, 3 H, including PhCH=CHCO, d, J = 15, at 6.89); 6.84-6.78 (2d, J = 8, 2 arom. H); 4.84-4.74 (m, PhCHN); 3.8-3.1 (m, 13 H); 2.85 (d, J = 17, 1 H); 2.45-2.35 (m, 1 H); 2.18-2.07 (m, 1 H); 2.08-1.83 (m, 6 H). ESI-MS: 495 ([M + H] $^+$).

[1,1,2,2,3,3,4,4- 2 H₈]Butane-1,4-diyl Bis(methanesulfonate) (13). According to [14], to a soln. of 0.857 g (8.7 mmol) of [1,1,2,2,3,3,4,4- 2 H₈]butane-1,4-diol (12) in CH₂Cl₂ (30 ml) and Et₃N (3 ml, 22 mmol), MsCl (2.27 g, 20 mmol) was added dropwise at 0° over a period of 5 min. The mixture was stirred for 1.5 h at r.t. and then evaporated to dryness *in vacuo* (50°). The crystalline residue was washed with H₂O and then with MeOH. Recrystallization from EtOH gave 1.6 g (100%) of colorless crystalline 13. M.p. 113°. TLC (AcOEt): R_f 0.8. 1 H-NMR ((D₆)acetone): 3.1 (s, 6 H).

(-)-(8S)-8-(4-Methoxyphenyl)-1,13-(4-tolylsulfonyl)[14,14,15,15,16,16,17,17- 2 H $_8$]-1,5,9,13-tetraazacycloheptadecan-6-one (14). To a soln. of (S)-11 (600 mg, 0.9 mmol) in DMF (95 ml), Cs $_2$ CO $_3$ (764 mg, 2.4 mmol) was added. The mixture was heated at 62° for 30 min. The soln. was cooled to r.t., and a soln. of 13 (248 mg, 1.3 mmol) in DMF (19 ml) was added dropwise. The mixture was stirred for 48 h at 55° and evaporated *in vacuo*. The residue was dissolved in 10 ml of H $_2$ O and extracted with CHCl $_3$ ($5 \times$), the org. layer was washed with H $_2$ O, dried (Na $_2$ SO $_4$), and evaporated. Compound (S)-14 was purified by CC (consecutively AcOEt, CHCl $_3$ /MeOH 20:1, CHCl $_3$ /MeOH 10:1): (S)-14 (353 mg, 54%). Colorless, glass-like solid. TLC (CHCl $_3$ /MeOH 10:1):

 R_1 0.61. [a] $_{10}^{10}$ = -15.4 (c = 3.25, CHCl₃). ¹H-NMR (CDCl₃): 8.24 (t, CONH); 7.68, 7.63 (2d, J = 8, 4 arom. H); 7.3 (t, J = 8, 4 arom. H); 7.15 (d, J = 9, 2 arom. H); 6.88 (d, J = 9, 2 arom. H); 3.89, 3.87 (2d, J = 9, PhCHN); 3.8 (s, MeO); 3.49 - 3.43 (m, 1 H); 3.39 - 3.01 (m, 5 H); 2.61 - 2.42 (m, 10 H, including 2 Me at 2.43, 2.42); 2.04 - 1.74 (m, 3 H); 1.72 - 1.65 (m, 2 H). ¹³C-NMR (CDCl₃): 172.8 (C=O); 162.4, 158.6, 143.2, 143.1, 135.7, 134.8, 129.6, 129.5, 127.3, 126.9, 113.9 (C=C); 58.8 (PhCN); 55.1 (MeO); 47.8, 47.3, 44.1, 43.8, 37.2, 36.6 (Me), 36.3, 31.2, 29.9, 29.5, 21.3 (CH₂). ESI-MS: 679 ([M + H] $^+$).

(-)-(8S)-(4-Methoxyphenyl)[14,14,15,15,16,16,17,17- 2H_8]-1,5,9,13-tetraazacycloheptadecan-6-one (15). The detosylation of (*S*)-14 was achieved by electrolysis according to [11]. The mixture was evaporated *in vacuo*. The residue was dissolved in 10 ml of H_2O , and the soln. was saturated with K_2CO_3 and extracted with a mixture of CHCl₃i-PrOH 4:1. The combined org. extracts were evaporated. The residue was dissolved in CHCl₃ and purified by CC (consecutively CHCl₃, CHCl₃/MeOH 7:3, CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): (*S*)-15 (129.7 mg, 80%). Colorless, glass-like solid. TLC (CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): R_f 0.54. $[\alpha]_D^{30} = -33.4$ (c = 3.5, CHCl₃). 1 H-NMR (CDCl₃): 8.38 (br. s, CONH \cdots N); 72 (d, J = 9, 2 arom. H); 6.86 (d, J = 9, 2 arom. H); 3.95 (m, PhCHN); 3.8 (s, MeO); 3.5 –3.44 (m, 1 H); 3.33 –3.28 (m, 1 H); 2.84 –2.8 (m, 1 H); 2.76 –2.67 (m, 2 H); 2.61 –2.4 (m, 8 H); 1.75 –1.49 (m, 4 H). 13 C-NMR (CDCl₃): 171.4 (C=O); 158.5 (C(4')); 135.2 (C(1')); 127.6 (C(3'), C(5')); 113.8 (C(2'), C(6')); 59.5 (PhCN); 55.1 (MeO); 48.9, 47.8, 46.7, 45.5, 38.7, 28.3, 28.0 (CH₂). ESI-MS: 371 ([M + H] $^+$).

(+)-(2S)-2-(4-Methoxyphenyl)[10,10,11,11,12,12,13,13-2 2 H₈]-1,5,9,14-tetraazabicyclo[12.3.1]octadecan-4-one (**16**). To a soln. of (*S*)-**15** (130 mg, 0.35 mmol) in 5 ml of MeOH was added 37% aq. HCHO soln. (32 μl, 0.4 mmol). After several min, the mixture was evaporated. The residue was purified by CC (consecutively, MeOH, CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): (*S*)-**16** (123.5 mg, 92%). Colorless, glass-like solid. TLC (CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): R_f 0.7. [α] $_D^{20}$ = +2.48 (c = 1.295, CHCl₃). H-NMR (CDCl₃): 8.95 (br. s, CONH ··· N); 7.06 (d, J = 8, 2 arom. H); 6.85 (d, J = 8, 2 arom. H); 3.94 (br. d, J = 10, PhCHN); 3.73 (s, MeO); 3.62 – 3.29 (m, 3 H); 2.97 – 1.94 (m, 10 H); 1.78 – 1.39 (m, 4 H). 13 C-NMR (CDCl₃): 171.8 (C=O); 158.8 C(4')); 129.1 (C(3'), C(5')); 128.7 (C(1')); 113.2 (C(2'), C(6')); 63.8 (PhCN); 55.1 (MeO); 52.5, 48.2, 38.2, 24.6 (CH₂). ESI-MS: 383 ([M + H] $^+$).

(+)-(2S)-2-(4-Methoxyphenyl)-9-[(E)-3-(4-methoxyphenyl)prop-2-enoyl][10,10,11,11,12,12,13,13-2+ H_8]-1,5,9,14-tetraazabicyclo[12.3.1]octadecan-4-one (17). Compound (S)-16 (110 mg, 0.29 mmol) was dissolved in CH₂Cl₂ (2 ml), and the soln. was cooled to 0°. A soln. of (*E*)-3-(4-methoxyphenyl)propenoyl chloride (prepared according to [15]; 75 mg, 0.38 mmol) in 4 ml of CH₂Cl₂ and Et₃N (300 μl) were then added dropwise. The mixture was stirred at 0° for 2 h. Then, the reaction was quenched by adding 5 ml of H₂O and allowed to warm to r.t. The aq. layer was extracted with CHCl₃ (4×). The combined org. layers were washed with H₂O and evaporated. The residue was purified by CC (consecutively, AcOEt, AcOEt/MeOH 9:1): (*S*)-17 (101 mg, 65%). Colorless solid. TLC (AcOEt/MeOH 4:1: R_f 0.45. $[\alpha]_D^{30} = +1.67$ (c = 2.30, CHCl₃). ¹H-NMR (CDCl₃): 9.45 (br. s, CONH···N); 7.68 (d, J = 15, PhCH=CHCO); 7.48 – 7.46 (m, 2 arom. H); 7.03 – 6.91 (m, 2 arom. H); 6.88 – 6.85 (m, 4 arom. H); 6.68 (d, J = 15, PhCH=CHCO); 3.98 (br. d, J = 12, PhCHN); 3.84, 3.8 (2s, 2 MeO); 3.66 – 2.36 (m, 9 H); 2.04 – 1.43 (m, 7 H). ESI-MS: 543 (M + H]+).

(-)-(8S)-8-(4-Methoxyphenyl)-1-[(E)-3-(4-methoxyphenyl)prop-2-enoyl][14,14,15,15,16,16,17,17- 2 H₈]-1,5,9,13-tetraazacycloheptadecan-6-one (**18**). Compound (S)-**17** (46.1 mg, 0.085 mmol) was hydrolyzed in 1% aq. HCl soln. (2.5 ml) at 70° for 1 h in the presence of NH₂OH·HCl (124 mg, 1.8 mmol). The mixture was extracted CHCl₃ (2×), the H₂O layer was alkalinized with 25% aq. NH₃ soln. and extracted with CHCl₃ (4×). The org. extract was washed with H₂O, dried (Na₂SO₄), and evaporated to give (S)-**18** (44 mg, 98%). Colorless, glass-like solid. [α]_D²⁰ = -8.06 (c = 2.27, CHCl₃). ¹H-NMR (CDCl₃, mixture of conformers): 8.37 (br. s, 0.5 H, CONH··N); 7.68 (d, d = 15, PhCH=CHCO); 7.52 – 7.49 (m, 2 arom. H); 7.32 (br. s, 0.5 H, CONH); 7.11 – 7.08 (m, 2 arom. H); 6.95 – 6.84 (m, 3 arom. H, including PhCH=CHCO); 6.74 – 6.72 (m, 2 arom. H); 3.94 – 3.01 (m, 11 H, including PhCHN, br. d at 3.93; 2s, 2 MeO at 3.85, 3.75); 2.85 – 2.34 (m, 8 CH); 2.08 – 1.4 (m, 4 CH). ¹³C-NMR (CDCl₃): 172, 166 (2 C=O); 161, 159, 142.4, 129.3, 127.9, 127.6, 115, 114.1, 113.8 (C=C); 58.9 (PhCN); 55.2, 55.1 (MeO). ESI-MS: 531 ([m + H]⁺).

(+)-(8S)-8-(4-Hydroxyphenyl)-1-[(E)-3-(4-hydroxyphenyl)prop-2-enoyl][14,14,15,15,16,16,17,17- 2 H₈]-1,5,9,13-tetraazacycloheptadecan-6-one Dihydrochloride (19; 2 HCl). All steps were performed under red light to avoid $(E) \rightarrow (Z)$ photoisomerization. A soln. of (S)-18 (44 mg, 0.08 mmol) in dry CH₂Cl₂ (2 ml) was cooled in a dry ice/acetone bath to -78° . A lm CH₂Cl₂ soln. of BBr₃ (1.5 ml, 1.5 mmol) was added. The mixture was kept at -78° for 1 h, then the bath was removed, and the mixture was stirred for 16 h at r.t. The reaction was quenched by adding 3 ml of 90% aq. MeOH soln. and evaporated to dryness $in\ vacuo\ (50^\circ)$. The excess BBr₃ was removed by repetitive addition of MeOH sat. with gas. HCl and evaporation. The solid residue was washed several times with CHCl₃. The residue was then dissolved in CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:0.3 and purified by CC

(consecutively, CHCl₃, CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:0.3). The eluate was evaporated to dryness. The residual free base of compound (S)-19 was transformed immediately to its dihydrochloride by addition of MeOH sat. with gas. HCl and evaporated to dryness: (S)-19·2 HCl (46 mg, 92%). Colorless solid. TLC (CHCl₃/MeOH/25% aq. NH₃ soln. 7:3:1): R_1 0.5. [α] $_{10}^{20}$ = +5.19 (c = 1.79, MeOH). 1 H-NMR ((D_4)MeOH, mixture of conformers): 7.56 – 7.53 (m, 3 H, including 2d, PhCH=CHCO, at 7.55, 7.54, J = 15); 7.43 – 7.37 (m, 2 arom. H); 6.96, 6.92 (2d, PhCH=CHCO); 6.88 – 6.84 (m, 2 arom. H); 6.83 – 6.79 (m, 2 arom. H); 4.9, 4.89, 4.86, 4.85 (4d, J = 10, PhCHN); 3.82 – 3.6 (m, 1.5 H); 3.57 – 3.38 (m, 2.5 H); 3.29 – 3.05 (m, 5 H); 2.86 – 2.8 (m, 1 H); 2.52 – 2.4 (m, 1 H); 2.14 – 2.04 (m, 1 H); 2.04 – 1.88 (m, 2 H). 1 H-NMR (MeOH): 3.5 (br. s, 2 1 H); 3.18 (br. s, 2 1 H); 1.8 (br. s, 4 1 H). 1 C-NMR ((D_4)MeOH): 173.4, 169.7 (2 C=O); 160.5, 159.4, 144.5, 131.3, 131.1, 128.0, 125.9, 117.2, 116.6, 115.2 (C=C); 58.9 (PhCN). ESI-MS: 503 ([M + H] $^{+}$).

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